

Molecular Perspectives on the Bantu Expansion: A Synthesis*

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Abstract

The expansion of Bantu-speaking peoples over large parts of sub-Saharan Africa is still a matter of debate—not only with respect to the propelling force behind it and the route(s) taken, but, also, in terms of the question whether there actually was a demographic expansion of peoples, rather than just a cultural expansion involving the spread of languages and technologies. In this paper, we provide a critical review of the extant linguistic and molecular anthropological data on Africa and discuss the insights they provide concerning the expansion itself as well as the demographic processes involved in it. Contrary to some assumptions by historians and cultural anthropologists, the genetic data speak in favor of an actual movement of peoples during the expansion of the Bantu languages over Africa, rather than a spread through language and culture shift. Furthermore, the molecular data indicate that sociocultural practices such as patrilocality and possibly even polygyny played a role in shaping the genetic diversity of Bantu-speaking peoples. These sociocultural practices might explain why, in Africa, there is a correlation between Y-chromosomal (i.e., paternal) lineages and linguistic affiliation, but not between mtDNA (maternal) lineages and language.

Keywords

Bantu languages; Bantu expansion; molecular anthropology; genetics

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1. Introduction

The dispersal of the Bantu language family is a topic that has incited a large amount of research, both amongst specialists in African history as well as from scholars working in other disciplines. Setting aside the higher phylogenetic levels of which they themselves are part, the Bantu languages constitute by far Africa's largest language family, both in terms of number of speakers and languages as well as in terms of geographical spread. About one African in three speaks a Bantu language, and estimates of the total number of languages vary between 440 (Guthrie, 1971) and ca. 680 (Mann and Dalby, 1987), mainly depending on where one draws the line between a language and a dialect (Nurse and Philippson, 2003a). As regards geographical distribution, the overall majority of communities living south of a line from southern Nigeria in the west to southern Somalia in the east and as far as the Cape are Bantu-speaking (Nurse and Philippson, 2003a). In spite of this numerical and geographical superiority, the Bantu family is widely recognized as forming only a low-level subgroup of the Niger-Congo phylum, being at most a subbranch of a branch of the Benue-Congo node (Schadeberg, 2003; Williamson and Blench, 2000). Blench (2006: 126) tentatively links the expansion of the Niger-Congo phylum with the improving climate at the beginning of the Holocene, i.e. from 12,000–10,000 BP onward. The beginning of the Bantu dispersal is usually situated much later. Vansina (1995: 52) proposes 5,000 BP on the basis of glottochronology. Contrasting the relatively low linguistic diversity within Bantu with the much higher diversity within Niger-Congo, Blench (2006: 134, 136) prefers a later date, 4,000 BP. As discussed in Bostoen (2007), this approximate time depth of 5,000–4,000 BP correlates with the definitive breakthrough of Neolithic technologies, such as macrolithic tools, polishing, and pottery, in the archaeological record of the Grassfields region of Cameroon (Lavachery, 2001). Slightly younger, but closely related technological assemblages have been recovered from several sites further south, i.e. in Cameroon, Gabon, the two Congos and the Central African Republic. These date from the early second millennium BCE until the last centuries BCE, and the younger they are, the further south(east) they are generally located. This gradual Neolithic expansion is commonly associated with the Bantu language dispersal (de Maret, 1994–1995). The striking contrast between this relatively shallow time depth and the current widespread distribution of the language family raises the key question of the 'Bantu Problem' (Eggert, 2005), namely, which factors were involved in the relatively rapid spread of this group of closely related languages over a disproportionately large part of sub-Saharan Africa (Bostoen, 2007; Eggert, 2005; Ehret, 2001; Vansina, 1979; 1995).

Historical linguists, archaeologists, and historians have had a long-standing debate in search of answers to this question. While linguists made the classifications available from which possible migration patterns could be derived, archaeologists provided insights into the cultural processes that may have enabled the Bantu expansion. The prevailing synthesis emerging from this protracted multidisciplinary quest for the driving forces behind the Bantu expansion is a model which identifies the dispersal of Bantu languages and the spread of the Early Iron Age industrial complex, consisting of pottery, metallurgy, domesticated plants and animals as well as a sedentary way of life, in sub-equatorial Africa as results of the same historical event—the migration of Bantu-speaking farmers (Huffman, 1980; Huffman and Herbert, 1994–1995; Oliver, 1966; Phillipson, 1977; 1985; 1989; Vansina, 1995). This model of the Bantu expansion has been proposed in different versions and the contents of the cultural package that accompanied the language spread vary from one proposal to another. In most models, metallurgy or grain cultivation are still absent during the first expansions through the equatorial rainforest, and only join the package when the dispersal in sub-equatorial Africa starts (de Maret, 1994–1995; Ehret, 1974; Vansina, 1990; 2006). This package model has been severely criticized for its lack of critical evaluation of underlying concepts and methods and for its propensity to circular argumentation (Eggert, 2005; Möhlig, 1989). Another recurrent criticism of the package model of the Bantu expansion concerns the overemphasis on migration as the single historical process underlying the spread of the Bantu languages (Gramly, 1978; Lwanga-Lunyiigo, 1976; Robertson and Bradley, 2000; Schepartz, 1988; Vansina, 1995). Nonetheless, this remains the reference model, especially for non-specialists, not least thanks to its explanatory elegance and straightforwardness.

Over the past 20 years, the debate has been joined by molecular anthropologists supplying an entirely new body of evidence that can be applied to the Bantu question and similar problems of language dispersal. As we discuss in this paper, recent developments in the field of molecular genetics have opened pathways that have the potential of leading to significant new insights into the demographic processes underlying the Bantu expansion. At the same time, unfortunately, many geneticists have been industrious in adding momentum to historical scenarios flawed by circular reasoning, rather than challenging earlier theories. Where Bantu speakers are involved, the Bantu migration model is often relied on for the historical interpretation of the geographic distribution of genetic markers. It is obvious that the risk of lapsing into assumptions that simply reinforce what has been stated before, instead of testing the original hypotheses, is particularly high: genetic data serve to reconstruct the demographic dynamics that may have underlain the Bantu language dispersal, yet it is precisely this preconceived Bantu migration model that is relied

on to interpret the genetic data. Especially among proponents of the farming/language dispersal hypothesis, the Bantu expansion is seen as a textbook case of the concurrent dispersal of early agriculture, human genes, and languages (Bellwood and Renfrew, 2002; Renfrew, 1992; 2002). Diamond and Bellwood (2003: 598) express this vision in the plainest way possible when describing the Bantu language dispersal as one of the world's clearest examples of "expanding farmers bearing their own archaeologically visible culture, domesticates, skeletal types, genes, and languages."

Such bold claims are definitely not a reflection of the present state of knowledge. As regards the beginning and spread of agriculture in the Bantu area, Neumann (2005) convincingly demonstrates that independent material evidence from the archaeological record is very scarce, since unambiguous supporting data on crop remains or agricultural tools are largely missing. Concerning skeletal types, the Bantu expansion has been associated with the spread of the so-called 'Negro race' over central and southern Africa (Hiernaux, 1968; 1974). Such associations have justly been discarded, however, not only because it has become abundantly evident that there is no biological basis for distinguishing races amongst humans on the whole (Cartmill, 1998; Serre and Pääbo, 2004; Templeton, 1998), but also because, in the absence of written documents, archaeological remains do not allow any insights into the language spoken by the people whose remains are studied. Furthermore, there are no chronological or geographical grounds to link the distribution of alleged 'Negro' skeletal material with a particular language family (Lwanga-Lunyigo, 1976; Robertson and Bradley, 2000), nor do mtDNA data "support the clustering of sub-Saharan Africans into (pre-Holocene) geographical races" (Salas et al., 2002: 1106).

A frequent problem that besets molecular anthropological studies is the implicit assumption that the modern-day distribution of ethnolinguistic groups is identical to their prehistoric distribution. However, it should be clear that the modern-day Bantu languages, which are presumed to have originated from one shared ancestor about 4,000 to 5,000 years ago as mentioned above, must have undergone a large amount of internal and contact-induced change to produce the striking diversity extant today. Thus, no single modern-day Bantu language can be equated with any ancestral language. Furthermore, ethnolinguistic communities are often fluid entities, shifting their allegiance over time. In Africa, most individuals are multilingual and so there is no one-to-one correlation between linguistic and genetic affiliation, which poses a problem for the investigation of language dispersals using current affiliations. These problems might appear to make the attempt at elucidating the past a fruitless waste of time and effort. However, the fact still remains that the very closely related Bantu languages are spread over a wide area of sub-Saharan Africa, and

this geographic spread needs to be explained. While languages, culture and ethnic affiliation can change relatively rapidly, genetic turnover can only occur through immigration and replacement of the pre-existing populations, such as occurred in historical times in Tasmania. This relative temporal stability of genetic material enables molecular anthropologists to draw inferences about prehistoric demographic events, and these inferences can be integrated with archaeological and linguistic data to provide a more complete perspective on African history. Of course, as pointed out by MacEachern (2000), such inferences can only be as good as the data they are based on; flawed sample sets will lead to erroneous conclusions. Nevertheless, as shown by recent, carefully conducted studies (Quintana-Murci et al., 2008; Veeramah et al., 2010), the field has come a long way towards solving such issues.

In this interdisciplinary review paper we aim to provide a critical assessment of the available molecular genetic evidence for the Bantu migration and correlated demographic processes of interest to historical linguists. Since the Bantu language family is first of all a linguistic unit, we start by introducing the linguistic evidence for the spread of these languages.

2. Historical Classification(s) of the Bantu Languages and Hypotheses of Migration

The historical unity of the Bantu languages is widely recognized. Since Greenberg (1972), the region between southeastern Nigeria and western Cameroon has generally been accepted as the Bantu homeland. It is not only the zone of highest linguistic diversity within Bantu, but Bantu languages also meet there with several other subgroups of the larger Benue-Congo family; amongst others, their closest relatives known as Bantoid languages. For lack of indisputable innovations that delineate Bantu from these closest relatives, the demarcation of the Bantu area is problematic. It is impossible to assume a clear-cut split between ‘Narrow’ and ‘Wide’ Bantu languages, i.e., the Bantu languages as conventionally classified by Guthrie (1948; 1971) vs. the closely related ‘Bantoid’ languages from NW-Cameroon and SE-Nigeria. The two groups form a linguistic continuum rather than two clearly distinct branches of the Southern Bantoid subgroup of Benue-Congo, which is itself one of the main branches of the Niger-Congo phylum (cf. Blench, 2006: 120; Nurse and Philippson, 2003b).

As regards the internal classification of Bantu languages, most attempts have relied on lexicostatistics (Bastin, 1983; Bastin et al., 1979; 1999; Coupez et al., 1975; Heine, 1973; Heine et al., 1977; Henrici, 1973; Nurse and Philippson, 2003b). This method produces language trees based on the calculation of lexical distance between language pairs in terms of percentages of shared

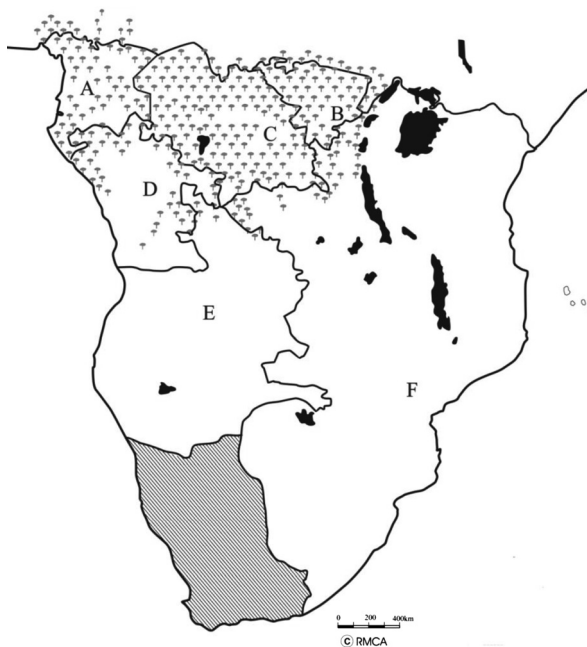


Figure 1. Approximate location of the major Bantu subgroups discussed in the text: A North-West, B Lebonya-Boan, C Inner Congo Basin, D West-Coastal, E South-West, F East.

cognates (cf. Nurse, 1997). Bantu classifications based on the classical comparative method, that is, on phonological or morphological innovatory features, are far less common and certainly do not result in the straightforward genealogical trees generated by lexicostatistics, due to the convergence effects to which Bantu languages have been exposed (Bastin, 1980; 1983; Ehret, 1999; Hyman, 1999; Möhlig, 1977; 1981; Nurse and Philippson, 2003b). Multilingualism as well as intensive and long-term contacts between Bantu speech communities have resulted in the transfer of features between closely related languages, which is often difficult to detect if it happened a long time ago. These convergence phenomena tend to blur linguistic frontiers and to complicate the demarcation of clear-cut Bantu subgroups. In addition, the internal relationships that exist between the subgroups and the way Bantu languages spread over their current territory are also a matter of debate. In contrast to what might be assumed, it is important to realize that no global proposal of internal Bantu language classification has yet received unanimous approval (Schadeberg, 2003).

It is generally acknowledged that there is considerably more historical diversity in the western part of the Bantu domain than in the eastern part. Bantu lan-

guages in the northwest, mostly in Cameroon and to a lesser extent in Gabon, are in many respects distinct from the other Narrow Bantu languages. Not only do they consistently turn up as a separate clade in lexicostatistical studies, they are also quite different typologically, e.g., in terms of maximality constraints on the verb stem (Hyman, 2004). For these reasons and because of the geographical proximity to the assumed Bantu homeland, they are generally considered as descendants of early breakaways from the Bantu nucleus (Bastin and Piron, 1999; Nurse and Philippson, 2003b; Vansina, 1995). Apart from a couple of isolated languages in the northeastern part of the DRC (the so-called Lebonya/Boan Bantu languages), the other western Bantu subgroups, i.e. Inner Congo Basin, West-Coastal Bantu and Southwest Bantu, are considered more closely related amongst each other than with the rest of Bantu languages and possibly go back to an intermediate common ancestor, sometimes labeled ‘Narrow West Bantu’ (Vansina, 1995). East Africa contains only the East Bantu subgroup, which is widely accepted as forming a coherent unit consisting of a large northeastern Savanna group in the north and several smaller groups in the south (Nurse and Philippson, 2003b: 175; see Fig. 1).

A major debate in historical Bantu studies concerns the question whether East Bantu is a primary Bantu branch or rather a later offshoot from a western Bantu node. This question is of great importance for our understanding of the Bantu language dispersal, seeing that two models of evolution prevail at present (Bostoen, 2004; Bostoen and Grégoire, 2007; Wiesmüller, 1997). According to the ‘East separate from West’ model, East Bantu is a primary branch of the Proto-Bantu node and its emergence in eastern Africa is the result of an eastward dispersion from Cameroon along the borders of the rainforest to the Great Lakes region. The spread of western Bantu is an entirely distinct event, characterized by a north-south movement through the equatorial rainforest and a rapid disintegration into different subgroups (Bastin et al., 1999; Coupez et al., 1975; Möhlig, 1981). In contrast, according to the ‘East out of West’ model, East Bantu is only a relatively late Bantu offshoot and constitutes a greater subgroup called ‘Savanna Bantu’ together with, roughly, the West-Coastal Bantu and Southwest Bantu subgroups, as opposed to the remainder of the Bantu languages, the so-called ‘Forest Bantu’ languages (Ehret, 2001; Heine et al., 1977; Henrici, 1973; Holden et al., 2005; Rexová et al., 2006; see Fig. 2). Roughly speaking, this latter scenario assumes an initial spread from the Bantu homeland through the tropical forest into the lower Congo region, from where a second wave would have started in different directions. One of these secondary migrations would be the origin of East Bantu. Several scholars, however, have argued against the historical validity of the so-called ‘Forest’ and ‘Savanna’ subgroups. Grégoire (2003) demonstrates that the forest is an area of great linguistic diversity, and Bantu languages spoken there do not

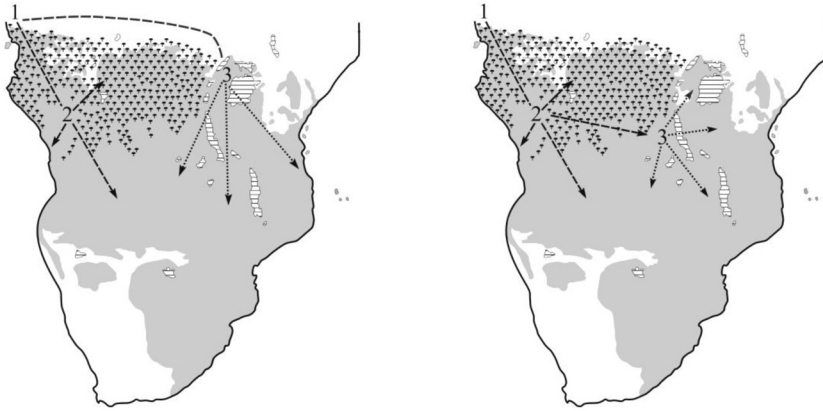


Figure 2. The two prevailing models of the Bantu expansion. 1 = Proto-Bantu nucleus, 2 = West-Bantu nucleus, 3 = East-Bantu nucleus. A: The ‘East separate from West’ model; B: The ‘East out of West’ model.

constitute a homogeneous subgroup that can be opposed to Savanna Bantu. Nurse and Philippson (2003b) have shown that several ‘Savanna’ languages from the western part of the Bantu range share several innovations with the ‘Forest’ languages to their north, which they do not have in common with the other ‘Savanna’ languages from the East. The current closeness between certain Savanna Bantu languages originally belonging to distinct eastern and western subgroups is most likely the result of sustained language contact involving profound convergence effects (see, for instance, Holden and Gray, 2006). As will be discussed in the following (cf. section 4.2), these contacts have also leveled the genetic differentiation between Bantu-speaking peoples in western and eastern Africa.

3. Molecular Anthropological Studies in Africa¹

Like other biological anthropologists, molecular anthropologists study the differences (polymorphisms) between humans, focusing, however, not on the phenotypic level such as hair or skin color, but on differences at the molecular level. Under the assumption of a single origin of modern humans, similarities in the genetic make-up of individuals or populations may indicate

¹ For a basic introduction to molecular anthropology, readers are referred to Appendix 1 in Pakendorf (2007), as well as to the following specialist introductory literature: for the Y chromosome, Jobling and Tyler-Smith (2003); for mtDNA, Pakendorf and Stoneking (2005); as well as to the textbook by Jobling et al. (2004).

either shared ancestry or subsequent admixture, while differences can indicate that they have had separate histories. It should be noted that a ‘population’ in molecular anthropological studies is not a given, hard-and-fast concept. While a population in theoretical population genetics is a unit of randomly mating individuals, this does not necessarily hold for molecular anthropological studies, where the units under investigation are often defined by the research questions. In general, attempts are made at defining the populations of interest based on linguistic and geographic information as well as self-identification. However, the scale may differ widely, depending on the interest of the investigators (cf. MacEachern, 2000). Thus, a study dealing with fine-scaled differences between subgroups within one or two ethnolinguistic groups (e.g., Pakendorf et al., 2007) may analyze the genetic variation in units corresponding to one or two settlements, while investigations of broad-scale geographic differentiation (e.g., Lao et al., 2008; Novembre et al., 2008) may analyze populations at the state level. These divergent levels of analysis do not in themselves constitute a problem as long as it is kept in mind what the units of investigation are, and as long as similar units are compared with each other. Problems can and do arise, however, when data from smaller ethnolinguistic communities are taken to represent larger geographic or linguistic groups, without the inherent limitations being kept in mind. Thus, Tishkoff et al. (2009) use two small samples of ‘South African Khoisan’ (six Ju speakers and eight !Xun/Khoe, i.e. a mixed group) to represent the ‘South African Khoisan’ groups as a whole; rather than being aware of the fact that these small samples cannot possibly be representative of all Khoisan-speaking groups from southern Africa, they make questionable claims about African prehistory on this basis.

Overall, the African continent is still severely understudied from a molecular anthropological perspective. The recent increase in molecular anthropological studies in Africa notwithstanding, large areas, especially in central and southern Africa, still remain blank, and among the studies that do exist, very few have undertaken to analyze both mitochondrial DNA (mtDNA) and Y-chromosomal variation in the same population groups,² thus leading to somewhat one-sided conclusions. MtDNA has two major advantages for studies of population history: first of all, it is present in the cell in high numbers of largely identical molecules that do not undergo recombination. Therefore, mutations remain within the physical sequence of nucleotides in which they first arose, and later mutations occur on the background of earlier ones; this permits the reconstruction of evolutionary trees of mutations. Secondly, mtDNA is inherited solely in the maternal line, thus shedding light on the maternal (i.e. female)

² Some exceptions are: Beleza et al. (2005); Destro-Bisol et al. (2004b); de Filippo et al. (2010); Knight et al. (2003); Tishkoff et al. (2007); see also the Appendix.

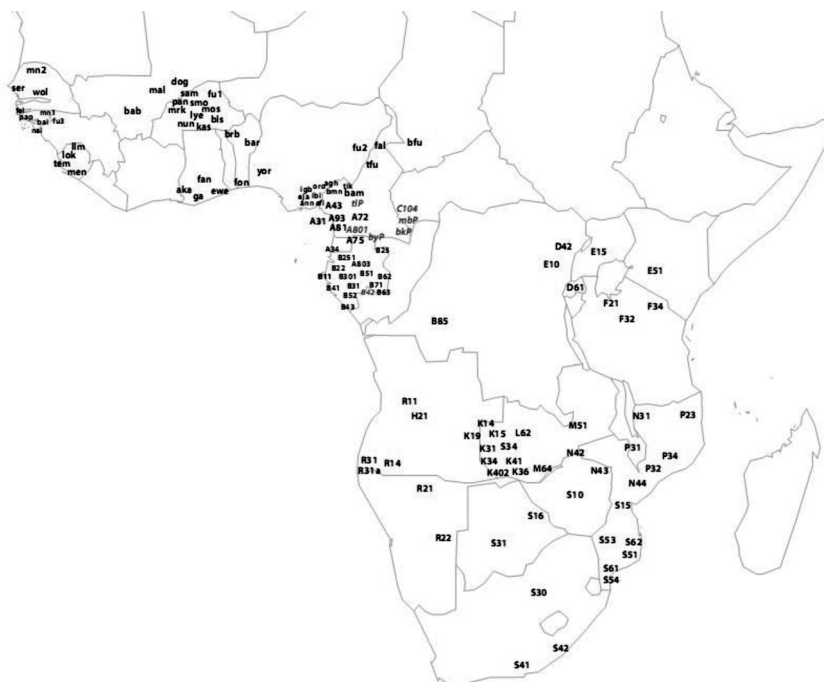


Figure 3. Map showing the approximate location of Niger-Congo samples included to date in molecular anthropological studies.

prehistory of the population. The counterpart of mtDNA is the Y chromosome, which has been the focus of numerous molecular anthropological studies in recent years. This chromosome, which determines the male sex of its carrier, is present only in men and is thus inherited only in the paternal line. Therefore, it illuminates the paternal history of a population. Like mtDNA, most of the Y chromosome does not undergo recombination,³ since there is only one such chromosome present in each cell. This permits the reconstruction of evolutionary trees for mutations arising on the Y chromosome, and therefore mutations can be traced back in time.

A relatively large number of populations speaking languages classified as Narrow Bantu have been included in diverse studies of African genetic variation (cf. the Appendix, incl. references therein); however, geographic coverage is very patchy, with eastern Africa, southwestern Africa, and central Africa particularly poorly covered (see Fig. 3).

³ In contrast to the mtDNA, however, which does not undergo recombination at all, a small part of the Y chromosome does recombine with the X chromosome.

Thanks to two recent studies of mtDNA and Y-chromosomal variation on the same individuals from Gabon and Cameroon (Quintana-Murci et al., 2008; Berniell-Lee et al., 2009), a large number of Northwest Bantu and West Coastal Bantu groups have now been analyzed. Eastern Africa is severely underrepresented with respect to Bantu-speaking groups, notwithstanding the importance of this region for investigating the hypotheses on the direction of Bantu migration. Only eight different Bantu-speaking ethnolinguistic groups have been included in molecular anthropological studies with sample sizes of 15 or more individuals, mostly with a focus on Y-chromosomal variation. Finally, southern Africa is by no means better represented, with the exception of Mozambique, from where several populations have been included in studies (Pereira et al., 2001;⁴ 2002; Salas et al., 2002; Sikora et al., 2011). For south African Bantu-speaking groups, mtDNA data are practically nonexistent, with a couple of exceptions. One study, which focused on a specific deletion on the mtDNA, sequenced the so-called hypervariable region in only a subset of samples (Soodyall et al., 1996), while complete mitochondrial genome sequences are available for a handful of individuals in a worldwide comparison (Kivisild et al., 2006). On the other hand, several groups from this region, such as Zulu, Xhosa, Sotho, and Tswana, have been analyzed for Y-chromosomal markers (Wood et al., 2005).

In recent years, large-scale studies of the variation present in the so-called autosomes (non-sex chromosomes, i.e. all the chromosomes excluding the X and Y chromosomes) have grown in importance. The autosomes have the advantage that they represent the bulk of the human genome, thereby providing a large amount of polymorphisms that can be analyzed. However, the autosomes are inherited from both parents and undergo recombination; it is therefore impossible to trace individual lineages back in time. Thanks to the development of innovative computational methods, however, such large-scale autosomal data can now be used for the investigation of demographic events, such as changes in population size, as well as proportions of shared ancestry—be it due to inheritance or to admixture between groups. The most important study of African genetic diversity to date was conducted by Tishkoff et al. (2009), who studied over 1,000 autosomal markers in over 2,000 African samples from 121 populations speaking languages belonging to various families (and including some ‘mixed populations,’ such as the !Xun/Khoe, mentioned above, or the Tutsi/Hutu). However, here, too, coverage is patchy, with a relatively small number of Bantu samples coming predominantly from Cameroon, Tanzania and Kenya, and a dearth of data from central and southern Africa.

⁴ This study, however, does not provide ethnolinguistic sample information.

This summary of molecular anthropological analyses of Bantu-speaking populations available to date has shown that there still exist large lacunae (cf. Fig. 3), especially with respect to populations from east, central, and southern Africa. Furthermore, the lack of data from both the Y chromosome and mtDNA makes it very difficult to come to comprehensive conclusions concerning the demographic processes involved in the Bantu expansion. It is therefore rather surprising that several papers refer to the Bantu expansion in their discussion or conclusions sections, generally claiming that the data presented support the assumption of such an event. As pointed out above, very few papers have undertaken a detailed study of the question, and in general the claims about the expansion are based on very scanty data and should be regarded with due caution. Nevertheless, as will be discussed in the following, even with the restricted amount of data currently available, it is possible to draw some conclusions regarding the relationships and history of Bantu-speaking populations.

4. Molecular Anthropological Perspectives on the Bantu Expansion

4.1. Correlations between Linguistic and Genetic Relationships

If one assumes a correlation between ‘genes’ and languages, then the classification of languages should reflect the genetic relationship of the groups speaking these languages to a certain extent. Such a correlation might be expected because, in general, children grow up learning the language of their parents, so that the transmission of genes and languages follows the same path. However, there are several factors that can confound such a correlation. Since humans can acquire any language, children may grow up speaking not the language of their parents, but that of the majority group surrounding them. Furthermore, human groups do not live in a vacuum, but come into contact with each other, which can have both genetic and linguistic effects. Population contact can lead to the interchange of genetic material through genetic admixture (intermarriage), confounding the regularity of correlations between the genetic and linguistic relationships of the groups concerned. Likewise, contact can lead to the interchange of linguistic material, be it copies of lexical or morphological items or copies of structural patterns. The most severe disruption of the correlation between genes and languages takes place when an entire ethnolinguistic group gives up its ancestral language and shifts to an unrelated language. However, although the effects of contact (which have certainly been at play in the Bantu languages, cf. section 2) can disrupt the correlation between the genetic and linguistic affiliation of a group on an individual scale, this connection might still hold at a broader level of comparison.

The correlation between linguistic and genetic affiliation in Africa was addressed by Scozzari et al. (1999) on the basis of Y-chromosomal data, Salas et al. (2002) on the basis of mtDNA data, and Wood et al. (2005) with a comparison of Y-chromosomal and mtDNA data. Scozzari et al. (1999) performed a very basic analysis of the amount of genetic variation found within populations grouped together by affiliation to one of the four Greenbergian language phyla (Afro-Asiatic, Niger-Congo, Nilo-Saharan, and Khoisan) as compared to the genetic variation found in large geographical groupings. They come to the conclusion that, overall, geography plays a larger role in structuring Y-chromosomal variation in Africa than linguistic affiliation does. A similar lack of correlation between genetic variation and linguistic affiliation was also found by Salas et al. (2002), who conclude that geography plays a larger role than linguistic affiliation in shaping mitochondrial DNA diversity in Africa. These results can be interpreted as evidence that marriage partners are chosen from geographically neighboring groups, irrespective of their linguistic affiliation—not entirely surprising, given the amount of multilingualism in Africa. In contrast, a very similar analysis by Wood et al. (2005), comparing Y-chromosomal with mtDNA data, leads to the result that while mtDNA variation does not correlate with linguistic affiliation, Y-chromosomal variation is better explained by linguistic affiliation to the four Greenbergian phyla than by geographical grouping. This finding is also supported by a finer-scale analysis performed on the basis of coarsely defined linguistic distances, suggesting “that African languages tend to be passed from father to children” (Wood et al., 2005: 873). In a recent study (de Filippo et al., 2011), we find a similar correlation between linguistic affiliation to the four major phyla and Y-chromosomal distances; however, the effects of geographical proximity (i.e., contact) are clearly discernible as well. This discrepancy between the correlation of linguistic affiliation with Y-chromosomal and mtDNA variation, respectively, can also be interpreted as evidence for the overall patrilocal social structure of African populations, in which brides are expected to move to their husbands’ place of residence. In such a case, even if the mothers of children come from a group speaking a different language, the children are surrounded by people speaking the same language as their fathers and grow up acquiring this language as their primary one.

Interestingly, while this correlation between linguistic affiliation and Y-chromosomal variation holds at the level of the four major linguistic phyla, it does not hold at a lower level. Thus, in a fine-scale study of genetic variation in ethnolinguistic groups speaking languages classified as Cross River, Southern Bantoid, and Igbo from the Cross River region in southeastern Nigeria, Veeramah et al. (2010) find that there is no correlation between either mtDNA or Y-chromosomal variation and linguistic affiliation; instead, there is a high level

of intermarriage between the groups. These results demonstrate that, at a small geographic scale, linguistic differences do not necessarily constitute a barrier to intermarriage, in good accordance with the high levels of multilingualism prevalent in Africa. Finer-scale molecular anthropological studies in different regions of Africa are necessary to obtain a better perspective on the degree of intermarriage and corresponding processes of language contact that have been at play in this continent.

4.2. *Genetic Markers and Evidence for the Bantu Migration*

One important factor that can shape the patterns of genetic variation in current-day populations is the fate of the pre-existing variation in the ancestral population. When a population splits into two or more daughter populations, each descendant group will receive some of the variation found in the ancestor. Depending on the geographic and cultural circumstances involved, the daughter populations may each receive a slightly different subset of the initial variation, leading to differences between them. Furthermore, random factors will cause the disappearance of some of the mutations that may initially have been present in the daughter populations. For instance, individuals carrying these polymorphisms might not reproduce, resulting in their genetic lineage dying with them, or men might have only daughters, or women might have only sons, resulting in the termination of their Y-chromosomal and mtDNA lineages, respectively. Such random processes affecting the genetic variation in a population are known as genetic drift. Genetic drift has a stronger effect in a small population than in a large one; thus, demographic factors such as population size play an important role in shaping genetic variation as well. An extreme case of genetic drift is the so-called founder event, in which a small group of individuals leaves the ancestral population and migrates to a different location. This founding group will carry only a small subset of the variation present in the entire ancestral population, and this subset will be present at high frequency in the resulting new population.

Thus, a notable reduction in genetic diversity can indicate prehistoric migration of a small group of founders. Long-distance expansions are expected to lead to serial founder events, with the genetic variation decreasing the further one moves from the point of origin, as seen in the expansion of modern humans out of Africa (Ramachandran et al., 2005). Should the expansion of the Bantu languages have involved an actual migration of people speaking these languages, we might expect to find signatures of serial founder events when moving further from the putative homeland in the Grasslands region of Cameroon. A cultural diffusion of the languages and agricultural technology without concomitant movement of people, on the other hand, would not

result in such a reduction of genetic diversity. It is therefore noteworthy that modern-day Bantu-speaking groups are characterized by low Y-chromosomal diversity. Only two haplogroups are found in the majority of ethnolinguistic groups: E1b1a* and its subgroup E1b1a7.⁵ A haplogroup is a group of related DNA molecules that is defined on the basis of shared mutations. This is similar to the linguistic innovations that are used by historical linguists to define nodes in trees depicting genealogical relationships between languages. The mutations defining specific haplogroups arise in a linear manner at different points in time on certain DNA molecules and spread through the population by reproduction; therefore, individuals carrying the same Y-chromosomal or mtDNA haplogroup are assumed to share a genetic ancestor. The distribution of particular haplogroups amongst communities speaking certain languages may thus be indicative of close genetic ties between these communities. However, there is no a priori evidence that would allow researchers to distinguish between sharing of haplogroups due to shared ancestry vs. genetic admixture (intermarriage).

The Y-chromosomal haplogroups E1b1a* and E1b1a7 account for 50–100 % of the Y-chromosomal variation in Bantu-speaking populations studied to date (Wood et al., 2005). E1b1a* is found in approximately 43 % of Bantu-speaking men, with a range from 17 % in the Hema to 66 % in the Ewondo. E1b1a7, on the other hand, is found on average in 30 % of Bantu-speaking men, with a range from 11 % in the Hema to 67 % in the Nande (Beleza et al., 2005; Cruciani et al., 2002; Knight et al., 2003; Wood et al., 2005). Even though these markers appear to be ‘Bantu-specific’ and have frequently been associated with the Bantu expansion (e.g. Berniell-Lee et al., 2009; Cruciani et al., 2002; Thomas et al., 2000; Zhivotovsky et al., 2004), neither of them is restricted in its distribution to Bantu-speaking populations;⁶ it should be noted at this point, however, that genetic markers need not be restricted to only one group in order to be informative about that group’s prehistory. As a matter of fact, markers are very rarely restricted to individual populations: a group may be characterized by high frequencies of a marker also found in other populations, especially if there was a founder event in their prehistory.

E1b1a* is found in very high frequency in west Africa in groups speaking languages classified as belonging to the Mande, Atlantic, Dogon, Gur, Kwa and Benue-Congo branches of Niger-Congo, such as Mandinka (79–82 %), Wolof (68 %), Mossi (67 %), Ewe (73 %), and others (Cruciani et al., 2002; Rosa et al., 2007; Wood et al., 2005); it is also found in non-Niger-Congo-speaking

⁵ As a matter of fact, de Filippo et al. (2011) show that the two major haplogroups found amongst Bantu speakers are actually subgroups of E1b1a* and E1b1a7, called E1b1a8 and E1b1a7a, respectively.

⁶ Cf. Appendix in Wood et al. (2005).

groups (Afro-Asiatic, Nilo-Saharan, and Khoisan). Although in some cases its presence in these communities might be due to admixture from Niger-Congo groups, the prevalence in Africa overall is so widespread that one cannot classify it as a Bantu-specific marker, nor as a marker characteristic of Niger-Congo speaking groups (cf. Quintana-Murci et al., 2010).

E1b1a7, on the other hand, which is a sub-lineage of E1b1a*, although not restricted to Bantu-speaking groups, does have a fairly restricted distribution in Africa. It is found predominantly in communities speaking languages of the Kwa and Benue-Congo branches of the Niger-Congo phylum, with the few exceptions being groups that may have obtained it through admixture with Bantu-speaking groups (e.g. the Mbuti pygmies or the Sekele !Kung; Cruciani et al., 2002; Wood et al., 2005). Thus, the distribution of this haplogroup in African populations provides some indication that the expansion of the Bantu-speaking groups over sub-Saharan Africa may have involved actual migration, not just the diffusion of languages and technology.

However, the genetic diversity associated with the sub-haplogroups E1b1a8 and E1b1a7a mentioned in footnote 5 does not diminish with distance from the putative homeland. This can be established with analyses of a different type of polymorphism, called short tandem repeats (STRs, also known as microsatellites). As indicated by the name, STRs are stretches of short repeated sequences between two and six base pairs long. DNA repetitions like these pose problems for the reproductive apparatus of the cell, leading to a high error rate in replication. Because of the high error (= mutation) rate of STRs, these are in general highly variable between individuals, permitting the investigation of the diversity present within haplogroups. Because the STR diversity within sub-haplogroups E1b1a8 and E1b1a7a does not diminish with geographic distance from the Bantu homeland, there is no evidence of the serial founder event one might expect if groups of people moved progressively through sub-Saharan Africa (de Filippo et al., 2011). This lack of evidence for migration might be interpreted as supporting the hypothesis of cultural diffusion, rather than actual migration of peoples, after all. What contradicts this suggestion is that there are no differences between Bantu-speaking groups in eastern Africa and those in western and central Africa with respect to the STR variation within these sub-haplogroups. This sharing of STR types between geographically distant Bantu groups would not be expected if they were originally indigenous groups who had only recently switched to the Bantu language and mode of subsistence. In such a case one would expect larger differences between western and eastern African Bantu-speaking groups, unless the shifting indigenous groups of western and eastern Africa were themselves closely related genetically—which, however, presupposes yet another recent spread of peoples over the continent, prior to the Bantu expansion. Thus, the

Y-chromosomal data do indicate actual migration of populations during the expansion of the Bantu languages. Combined with this evidence for actual movement of peoples, the lack of serial founder events indicates that either the migrating groups were large enough that there was no reduction in genetic diversity, or that the signal of an original founder event has subsequently been erased by later migrations—in good accordance with the linguistic evidence for contact (cf. section 2).

MtDNA diversity, on the other hand, is quite high in Bantu-speaking populations, even in those groups settled at the further end of the putative migration, such as Mozambique or Zambia (de Filippo et al., 2010; Salas et al., 2002). This high diversity can be interpreted as an indication that there were successive waves of genetically diverse migrations, or that the Bantu expansion involved a large group of migrants that took their origin in a diverse ancestral population. An alternative explanation for the relatively high mtDNA diversity in Bantu-speaking groups is that intermarriage with mostly women from indigenous communities took place in the course of the Bantu expansion, as will be discussed in more detail in section 4.3.2 below—that is, that the expansion involved predominantly migration of men.

Nevertheless, there is some indication that the dispersal of Bantu-speaking groups to southeastern Africa involved a founder effect. This is seen in the high frequency and reduced diversity of both mtDNA haplogroups L0a1a and L0a2 in the Mozambican Bantu-speaking groups (Salas et al., 2002: their Fig. 4) as well as haplogroups L2a1a and L2a1b (Salas et al., 2002: their Fig. 6). L2a1b in particular is interesting because it shows signs of expansion after the founder effect: a high-frequency haplotype from which a number of haplotypes that differ by only one mutation are derived (Salas et al., 2002: their Fig. 6). This is corroborated by the fact that the sequence diversity in Mozambique (0.96 ± 0.01) is slightly lower than that found in most west or east African populations. More investigations as detailed as that undertaken by Salas et al. (2002) are needed for other regions of Africa to elucidate the demographic factors at play during the course of the Bantu expansion.

In their study of autosomal variation, Tishkoff et al. (2009) find a strong signal of genetic relationship of the Niger-Congo groups throughout sub-Saharan Africa, possibly reflecting the spread of Bantu-speaking peoples. This signal is due to similar allele frequencies of the polymorphic markers investigated and not to the persistence of a specific lineage, such as the Y-chromosomal and mtDNA haplogroups discussed above. In contrast to the lack of differentiation within the Y-chromosomal sub-haplogroups E1b1a8 and E1b1a7a between western and eastern African Bantu-speaking groups, as discussed above, they also find weaker signals for “subtle substructure between East African

Bantu-speakers [...] and West-Central African Bantu-speakers [...], and individuals from Nigeria and further west, who speak various non-Bantu Niger-Kordofanian languages” (Tishkoff et al., 2009: 1039). Interestingly, in a recent study using ~2,800 autosomal markers in a set of 11 populations of African descent, Sikora et al. (2011) come to the conclusion that the expansion of Bantu languages to southeastern Africa (Mozambique) may well have taken place not so much through actual migration of peoples as through the assimilation of local groups who shifted to the Bantu language. The discrepancy between the findings of Tishkoff et al. (2009) and Sikora et al. (2011) might be due to the different populations included—notably, the study by Tishkoff et al. lacks Mozambican populations entirely, and populations from southern Africa as a whole are severely underrepresented. On the other hand, the study by Sikora et al. (2011) is based on a very restricted sample of Bantu-speaking groups: other than 180 samples from Mozambique, which are not ethnolinguistically defined, they include published data on two very small and undefined samples of ‘Bantu’ from Kenya and South Africa, as well as a sample of Luhya. These samples are clearly not representative of the genetic variation present in Bantu-speaking populations. Another factor that may explain the divergent results is the differences in markers analyzed by the two studies: Tishkoff et al. investigated a large number of highly variable STRs that play no functional role in the genome, while Sikora et al. studied slowly evolving sites in genes associated with immunity to diseases and inflammation response. The latter are expected to underlie selective pressures.

Although Salas et al. (2002) discuss their results in terms of the ‘East separate from West’ model (cf. section 2), while Plaza et al. (2004) suggest that the western and eastern streams of migration came together south of the rainforest and spread jointly throughout southern Africa, the preceding discussion should have made it clear that there is not yet sufficient data to warrant any conclusion concerning the potential route(s) of migration taken by the ancestors of modern Bantu-speaking groups. As described in section 3, there are still hardly any data from central, southwestern, eastern, or southern Africa. Thus, the genetic investigation of the putative routes of the Bantu expansion will have to be suspended until sufficient data have been collected. However, as mentioned above, the lack of genetic differentiation between Bantu-speaking groups from eastern Africa and central and western Africa with respect to the sub-haplogroups E1b1a8 and E1b1a7a may indicate that subsequent smaller migrations between Western and Eastern Bantu groups have erased the genetic signal of the initial split (de Filippo et al., 2011).

4.3. *Genetic Perspectives on Sociocultural Parameters Involved in the Bantu Dispersal*

4.3.1. Assimilation of Non-Bantu Speech Communities

A topic that has generated much interest among molecular anthropologists is the degree of admixture between the putative immigrating Bantu-speaking groups and the resident populations. Certain modern African hunter-gatherers are characterized by specific mtDNA and Y-chromosomal polymorphisms; therefore prehistoric admixture between incoming agriculturalists and resident hunter-gatherer groups genetically resembling modern foragers is detectable. For example, western pygmy groups are characterized by very high frequencies of mtDNA haplogroup L1c1a (~84%; Quintana-Murci et al., 2008), while southern African groups speaking Ju languages are characterized by haplogroups L0d and L0k. Similarly, the Y-chromosomal lineage A3b1 is characteristic of southern African Ju-speaking groups (Cruciani et al., 2002; Wood et al., 2005), while haplogroup B2b is more generally characteristic of African hunter-gatherer groups (Güldemann and Stoneking, 2008). However, as yet, relatively few hunter-gatherer groups have been included in molecular anthropological studies in Africa, so the prevalent idea of what would constitute ‘hunter-gatherer introgression’ is potentially biased and might lead to wrong conclusions.

From its distribution in current African populations, one could hypothesize that some mtDNA sub-clades of haplogroup L1c1a entered Bantu-speaking populations through admixture with central African pygmy groups: it is found in exceedingly high frequencies in western pygmies, as noted above, and is otherwise restricted to Bantu-speaking groups, with the exception of the Chadic-speaking Mandara (Coia et al., 2005) (5%) as well as the Yoruba, whose language is part of the Benue-Congo branch of Niger-Congo (6% of the sequences reported by Watson et al., 1996 and Vigilant et al., 1991). However, contrary to this hypothesis of admixture between the ancestors of the Bantu-speaking groups and the ancestors of modern-day pygmy groups, Quintana-Murci et al. (2008) propose that L1c was already present in the shared ancestor of agricultural groups (in their study represented by Bantu-speaking groups from Gabon and Cameroon) and pygmy groups; in addition, they suggest gene flow to have taken place over a long period of time after an extended period of initial isolation.

In southwestern Africa (Angola and the Angolan enclave Cabinda) there is no evidence for introgression of lineages characteristic of Ju-speaking groups (mitochondrial haplogroups L0d or L0k, Y-chromosomal A3b1 or B2b), which has been interpreted to indicate a replacement of local hunter-gatherers by

immigrating Bantu speakers (Beleza et al., 2005; Plaza et al., 2004). However, as mentioned above, our knowledge of the genetic composition of African hunter-gatherer groups is as yet very limited and such conclusions cannot be regarded as definitive. For instance, if the local hunter-gatherer groups had a genetic composition very different from the Ju-speaking !Kung groups used as the prototypical ‘Khoisan hunter-gatherers’ in molecular anthropological studies, but similar to that of the immigrating groups,⁷ admixture between the immigrating Bantu speakers and resident groups would not be detectable with the methods currently employed. Furthermore, since there is as yet no evidence that populations related to southern African Ju- or Tuu-speaking groups were formerly settled much further north than their current territory (Tom Güldemann, pers. comm.), lack of Ju-specific genetic markers in the immigrant Bantu communities does not necessarily provide evidence for a lack of admixture between the immigrants and local hunter-gatherers. On the other hand, the presence of clicks in some Bantu languages of southwestern Zambia demonstrates that some contact must have taken place in southwestern Africa (Bostoen and Sands, to appear).

In southeastern Africa, on the other hand, the Bantu-speaking groups appear to have intermarried to a minor degree with local hunter-gatherer groups that were genetically related to modern-day Ju-speakers, as evidenced by some level of ‘hunter-gatherer specific’ haplogroups (such as mtDNA L0d and Y-chromosomal A3b1) found in Bantu-speaking groups (de Filippo et al., 2010; Pereira et al., 2001; Salas et al., 2002; Wood et al., 2005). Interestingly, Southeast Bantu groups (Zulu, Xhosa, Sotho, and Tswana) have 28.5% of haplogroup L0d (Schlebusch et al., 2009); this is in good accordance with the linguistic facts, since the speakers of Zulu and Xhosa have adopted quite an extensive number of click phonemes into their languages (Güldemann and Stoneking, 2008: their Table 2). Thus, both the linguistic and the genetic data confirm that some of the Bantu-speaking groups who migrated to south Africa entered into close cultural and physical contact with the indigenous, click-using hunter-gatherers. However, while there is evidence of intermarriage with resident communities in southern Africa, the genetic data do not provide any indication that the spread of the Bantu languages and culture in this region was due only to shift of indigenous peoples. In such a case, we would expect to find a majority of forager-specific lineages in the Bantu-speaking groups, and not only a third or less. This, too, provides evidence that the Bantu expansion involved migration of people rather than just cultural diffusion, at least in this part of the continent.

⁷ Cf. the difference between the ‘Vasikela Kung’ and the Khwe (Chen et al., 2000).

The contact between Bantu-speaking groups and hunter-gatherers is suggested to have been largely sex-biased, with a higher degree of female migration from hunter-gatherer groups into agricultural groups, and a higher degree of male migration from agricultural groups to the hunter-gatherers. This is accounted for by the fact that it is considered unacceptable for women from agriculturalist communities to marry men from foraging communities, while the reverse is more acceptable. Thus, hunter-gatherer mtDNA lineages can enter agriculturalist communities; conversely, if such unions break up and the woman returns to her parental community with her children, any sons by her agriculturalist husband will be carrying his Y chromosome, which then gets incorporated into the gene pool of the foraging community (Cavalli-Sforza, 1986). This sex-biased admixture pattern between farmers and foragers has been confirmed by several studies (Destro-Bisol et al., 2004b; Quintana-Murci et al., 2008; Tishkoff et al., 2007; Wood et al., 2005). The fact that such socio-cultural boundaries to intermarriage are not watertight, however, is shown by the south African Sotho, Tswana, Zulu, and Xhosa, which have a frequency of 3–7% of the ‘Ju-specific’ Y-chromosomal haplogroup A3b1 (Wood et al., 2005). But since these groups have incorporated four to nine times as many hunter-gatherer female lineages (28.5% on average), as described above, here, too, the gene flow from indigenous groups to immigrating Bantu speech communities was predominantly female-mediated.

4.3.2. Demographic Features of the Bantu Migrations

Genetic variation on the mtDNA and Y chromosome is strongly shaped by cultural factors such as social organization, polygyny and marriage practices (Ségurel et al., 2008). In most of the worldwide populations studied by molecular anthropologists up to now, the genetic diversity of the Y chromosome is smaller than that of mtDNA, and the genetic differences between populations are larger when Y-chromosomal markers are analyzed than when mtDNA is used. One reason for this could be the differential reproductive success of men in polygynous societies, where a few men can afford to have several wives and thus father more children than the majority of men, who can only afford one wife, if any, and thus father fewer or no children (Hammer et al., 2008; Kayser et al., 2003). Another reason could be that the vast majority of human societies have patrilocal residence patterns so that the women (and with them their mtDNA) move to their husbands’ homes, while the men (and their Y chromosomes) stay in the village in which they were born (Oota et al., 2001; Seielstad et al., 1998). This leads to an incrementally higher amount of mixing of mtDNA than Y-chromosomal lineages, and results in higher mtDNA diversity.

Although it is difficult to compare values of mtDNA and Y-chromosomal diversity directly, since the underlying markers are ascertained in very different ways, there is still a general trend for mtDNA diversity amongst Bantu-speaking populations to be larger than the Y-chromosomal diversity (cf. section 4.2). As stated by Salas et al. (2002: 1105), most of the mtDNA haplogroups found in sub-Saharan Africa as a whole are found in the Bantu-speaking groups from Mozambique. On the other hand, all Bantu-speaking populations are characterized by very high frequencies of only two Y-chromosomal haplogroups, E1b1a* and its derivative E1b1a7⁸ (Wood et al., 2005). Furthermore, genetic differences between populations are larger when using Y-chromosomal markers than mtDNA, suggesting that the paternal lineages are mixing less than the maternal lineages. These discrepancies between mtDNA and Y-chromosomal variation might be indicative of sex-biased sociocultural practices such as patrilocality, which may have affected the demographics of men and women in different ways (Hammer et al., 2008; Kayser et al., 2003; Oota et al., 2001; Pilkington et al., 2008; Seielstad et al., 1998). As discussed above, the Y-chromosomal evidence speaks in favor of actual migration of peoples in the course of the expansion of Bantu languages and agricultural technology. The differences between Y-chromosomal and mtDNA variation in modern-day Bantu-speaking populations imply that, when the ancestors of the Bantu speakers migrated, they took in wives from the local groups but not husbands, following patrilocal marriage rules. In this way, the distinctive Y-chromosomal composition of the migrating Bantu speakers stayed relatively stable, while new mtDNA lineages were continuously added to the Bantu gene pool. This is also shown by the incorporation of more forager mtDNA lineages than Y chromosomes in south African Bantu groups, as discussed above.

It has been suggested that the paternal gene pool of the Bantu-speaking groups may further have remained quite small through the practice of polygyny (cf. Wood et al., 2005). A man who has several wives can expect to father more offspring than a man who has only one or no wife; therefore, over time the genetic lineages of the few men with more than one wife will increase in frequency. This, conversely, leads to a decrease in genetic diversity of the Y chromosome, since the same Y-chromosomal lineage is being passed on to proportionally more sons, who in turn will pass it on to a large number of grandsons, great-grandsons, and so forth. While the differences in Y-chromosomal and mtDNA diversity are not nearly as strong in Africa as they are in the highlands of western New Guinea (Kayser et al., 2003), the Y-chromosomal diversity tends to be lower, providing some indication that polygynous practices might

⁸⁾ Or, as shown by de Filippo et al. (2011), of the sub-haplogroups E1b1a8 and E1b1a7a.

have played a role. However, this hypothesis remains to be tested by comparing Y-chromosomal diversity values between polygynous and non-polygynous societies.

5. Conclusions

The above discussion of the extant molecular anthropological data on African populations, especially those speaking Bantu languages, has shown that sweeping conclusions concerning the nature of the ‘Bantu migration(s)’ are premature, and that current efforts at elucidating the demographic processes and patterns of migration are hampered by a severe shortage of data as well as the lack of comparability amongst them.

However, the current state of molecular anthropological studies on African populations indicates that the Bantu language dispersal actually involved human migration, not just the diffusion of languages and technology. Y-chromosomal diversity in Bantu-speaking populations is lower than mtDNA diversity, with only two Y-chromosomal haplogroups present in high frequency among most Bantu speech communities: E1b1a8 and E1b1a7a. Although neither of them is a Bantu-specific marker, and they are not even restricted to Niger-Congo-speaking populations, their current distribution in African populations suggests that male migrants carrying these haplogroups were involved in the spread of Bantu languages.

On the other hand, the higher mtDNA diversity among Bantu-speaking populations is indicative of sex-biased sociocultural customs such as patrilocality, whereby Bantu-speaking groups commonly intermarried with local women during their expansion, rather than incorporating indigenous men. The presence of characteristic central African ‘pygmy’ mtDNA haplogroups, such as L1c1a, and mtDNA haplogroups characteristic of southern African Ju-speaking groups, such as L0d, in the maternal gene pool of certain Bantu-speaking groups provides evidence of such female-mediated gene flow from local communities into migrating Bantu speech communities.

To be sure, until coverage of central, eastern and southern Africa has been ameliorated, these conclusions can only be viewed as provisional. Nevertheless, molecular anthropological studies, and especially recent revolutionary innovations in the available methods, have the potential of providing novel insights into the demographic processes that underlie the expansion of the Bantu languages over much of sub-Saharan Africa. Thus, novel techniques that investigate variation at a vast number of polymorphisms in the entire genome can now be coupled with simulation-based analyses of the data, which may provide more definitive insights into the genetic history of Bantu populations.

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Appendix. Ethnolinguistically defined Niger-Congo populations included in molecular anthropological studies to date, restricted to the uniparental markers (mtDNA and Y chromosome)

While this table is as accurate as possible, it is not always easy to trace samples through several publications to their original source; therefore, some errors may have slipped in, especially with respect to the sample numbers. Not all the studies referenced here as having included African samples deal primarily with the prehistory of African populations; in some cases, samples from African ethnolinguistic groups were included in studies of worldwide variation. On the other hand, some studies, such as the investigation of genetic variation in the Angolan enclave of Cabinda (Beleza et al., 2005), do not provide ethnolinguistic information on the samples included; these have been excluded from the table.

Populations with sample sizes smaller than 15 are omitted, with a few exceptions where the same population has more than 14 individuals typed either for the mtDNA or Y chromosome. In the column ‘subgroup,’ the language code according to Guthrie’s (1971) and Maho’s (2003) classification of Bantu languages is given in parentheses, while the column ‘ID’ contains the code used in Fig. 3. The column labeled ‘N’ gives the sample sizes for studies of mtDNA and Y chromosome, respectively. The references are included in the bibliography at the end of the paper.

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|------------------|----------|------------------|-------------------|---|--------------|--|
| | | | N | Study | N | Study |
| Baka Pygmies | Adamawa | bkP | 127 | Quintana-Murci et al., 2008 | 56 | Wood et al., 2005; Berniell-Lee et al., 2009 |
| Fali | Adamawa | fal | 40 | Černý et al., 2007 | 39 | Scozzari et al., 1997; Scozzari et al., 1999 |
| Balanta | Atlantic | bal | 62 | Rosa et al., 2004 | 26 | Rosa et al., 2007 |
| Borgor Fulani | Atlantic | bfu | 49 | Černý et al., 2007 | – | – |
| Felupe Ejamat | Atlantic | fel | 50 | Rosa et al., 2004 | 50 | Rosa et al., 2007 |
| Fulbe | Atlantic | fu1, fu2, fu3 | 61 | Vigilant et al., 1991; Watson et al., 1996; Kivisild et al., 2006 | 59 | Scozzari et al., 1997; Destro-Bisol et al., 2004b; Rosa et al., 2007 |
| Limba | Atlantic | lim | 67 | Jackson et al., 2005 | – | – |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|--------------------|-------------|-----|-------------------|---|--------------|--|
| | | | N | Study | N | Study |
| Nalu | Atlantic | nal | 26 | Rosa et al., 2004 | 17 | Rosa et al., 2007 |
| Papel | Atlantic | pap | 77 | Rosa et al., 2004 | 64 | Rosa et al., 2007 |
| Serer | Atlantic | ser | 23 | Rando et al., 1998 | – | – |
| Tcheboua Fulani | Atlantic | tfu | 40 | Černý et al., 2007 | – | – |
| Temne | Atlantic | tem | 121 | Jackson et al., 2005 | – | – |
| Wolof | Atlantic | wol | 48 | Rando et al., 1998 | 34 | Wood et al., 2005 |
| Annang | Cross-River | ann | 110 | Veeramah et al., 2010 | 110 | Veeramah et al., 2010 |
| Efik | Cross-River | efi | 148 | Veeramah et al., 2010 | 148 | Veeramah et al., 2010 |
| Ibibio | Cross-River | ibi | 516 | Veeramah et al., 2010 | 516 | Veeramah et al., 2010 |
| Oron | Cross-River | oro | 101 | Veeramah et al., 2010 | 101 | Veeramah et al., 2010 |
| Yoruba | Defoid | yor | 34 | Vigilant et al., 1991; Watson et al., 1996 | 13 | Tishkoff et al., 2007; de Filippo et al., 2011 |
| Dogon | Dogon | dog | – | – | 55 | Wood et al., 2005 |
| Kassena | Gur | kas | – | – | 33 | de Filippo et al., 2011 |
| Lyela | Gur | lye | – | – | 40 | de Filippo et al., 2011 |
| Mossi | Gur | mos | 10 | Kivisild et al., 2006 | 92 | Scozzari et al., 1997; Scozzari et al., 1999; de Filippo et al., 2011 |
| Nuna | Gur | nun | – | – | 29 | de Filippo et al., 2011 |
| Pana | Gur | pan | – | – | 19 | de Filippo et al., 2011 |
| Samoya | Gur | sam | – | – | 21 | de Filippo et al., 2011 |
| Ewe | Kwa | ewe | 88 | Veeramah et al., 2010 | 118 | Wood et al., 2005; Veeramah et al., 2010 |
| Fante | Kwa | fan | – | – | 32 | Wood et al., 2005 |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|------------------|-------------------------|-------------|-------------------|-----------------------------|--------------|---|
| | | | N | Study | N | Study |
| Fon | Kwa | fon | – | – | 36 | Scozzari et al., 1999 |
| Ga | Kwa | ga | – | – | 29 | Wood et al., 2005 |
| Akan | Kwa | aka | 155 | Veeramah et al., 2010 | 155 | Veeramah et al., 2010 |
| Bambara | Mande | bab | 52 | González et al., 2006 | – | – |
| Bissa | Mande | bis | – | – | 40 | de Filippo et al., 2011 |
| Loko | Mande | lok | 29 | Jackson et al., 2005 | – | – |
| Malinke | Mande | mal | 31 | González et al., 2006 | – | – |
| Mandenka | Mande | mn1, mn2 | 119 | Graven et al., 1995 | 99 | Wood et al., 2005; Rosa et al., 2007; de Filippo et al., 2011 |
| Marka | Mande | mrk | – | – | 33 | de Filippo et al., 2011 |
| Mende | Mande | men | 59 | Jackson et al., 2005 | – | – |
| Samo | Mande | smo | – | – | 78 | de Filippo et al., 2011 |
| Bariba | Voltaic | bar | – | – | 34 | Scozzari et al., 1999 |
| Berba | Voltaic | brb | – | – | 20 | Scozzari et al., 1999 |
| Igbo | West Benue- Congo | igb | 209 | Veeramah et al., 2010 | 209 | Veeramah et al., 2010 |
| Aghem | Bantoïd | agh | 118 | Veeramah et al., 2010 | 118 | Veeramah et al., 2010 |
| Bamileke | Bantoïd | bam | 48 | Destro-Bisol et al., 2004a | 48 | Scozzari et al., 1997; Caglià et al., 2003; Coia et al., 2004 |
| Bamun | Bantoïd | bmn | 117 | Veeramah et al., 2010 | 117 | Veeramah et al., 2010 |
| Ejagham | Bantoïd | eja | 138 | Veeramah et al., 2010 | 117 | Veeramah et al., 2010 |
| Tikar | Bantoïd | tik | 34 | Veeramah et al., 2010 | 34 | Veeramah et al., 2010 |
| Tikar Pygmies | Bantoïd | tiP | 35 | Quintana-Murci et al., 2008 | – | – |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|------------|--------------|------|-------------------|---|--------------|---|
| | | | N | Study | N | Study |
| Bubi | Bantu (A31) | A31 | 45 | Mateu et al., 1997 | – | – |
| Benga | Bantu (A34) | A34 | 50 | Quintana-Murci et al., 2008 | 48 | Berniell-Lee et al., 2009 |
| Bassa | Bantu (A43) | A43 | 47 | Destro-Bisol et al., 2004b | 49 | Destro-Bisol et al., 2004b |
| Ewondo | Bantu (A72) | A72 | 53 | Destro-Bisol et al., 2004a; Quintana-Murci et al., 2008 | 30 | Scozzari et al., 1997; Caglià et al., 2003; Coia et al., 2004 |
| Fang | Bantu (A75) | A75 | 105 | Quintana-Murci et al., 2008 | 64 | Berniell-Lee et al., 2009 |
| Makina | Bantu (A803) | A803 | 45 | Quintana-Murci et al., 2008 | 43 | Berniell-Lee et al., 2009 |
| Ngoumba | Bantu (A81) | A81 | 88 | Quintana-Murci et al., 2008 | 55 | Wood et al., 2005; Berniell-Lee et al., 2009 |
| Bakaka | Bantu (A93) | A93 | 50 | Destro-Bisol et al., 2004b | 49 | Destro-Bisol et al., 2004b |
| Galoa | Bantu (B11) | B11 | 51 | Quintana-Murci et al., 2008 | 47 | Berniell-Lee et al., 2009 |
| Orungu | Bantu (B11) | B11 | 20 | Quintana-Murci et al., 2008 | 21 | Berniell-Lee et al., 2009 |
| Akele | Bantu (B22) | B22 | 48 | Quintana-Murci et al., 2008 | 50 | Berniell-Lee et al., 2009 |
| Kota | Bantu (B25) | B25 | 56 | Quintana-Murci et al., 2008 | 53 | Berniell-Lee et al., 2009 |
| Shake | Bantu (B251) | B251 | 51 | Quintana-Murci et al., 2008 | 43 | Berniell-Lee et al., 2009 |
| Eviya | Bantu (B301) | B301 | 38 | Quintana-Murci et al., 2008 | 24 | Berniell-Lee et al., 2009 |
| Mitsogo | Bantu (B31) | B31 | 64 | Quintana-Murci et al., 2008 | 60 | Berniell-Lee et al., 2009 |
| Eshira | Bantu (B41) | B41 | 40 | Quintana-Murci et al., 2008 | 42 | Berniell-Lee et al., 2009 |
| Punu | Bantu (B43) | B43 | 52 | Quintana-Murci et al., 2008 | 58 | Berniell-Lee et al., 2009 |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|------------|---------------|-----|-------------------|--|--------------|--|
| | | | N | Study | N | Study |
| Duma | Bantu (B51) | B51 | 47 | Quintana-Murci et al., 2008 | 46 | Berniell-Lee et al., 2009 |
| Nzebi | Bantu (B52) | B52 | 63 | Quintana-Murci et al., 2008 | 57 | Berniell-Lee et al., 2009 |
| Obamba | Bantu (B62) | B62 | 47 | Quintana-Murci et al., 2008 | 47 | Berniell-Lee et al., 2009 |
| Ndumu | Bantu (B63) | B63 | 39 | Quintana-Murci et al., 2008 | 36 | Berniell-Lee et al., 2009 |
| Ateke | Bantu (B71) | B71 | 54 | Quintana-Murci et al., 2008 | 48 | Berniell-Lee et al., 2009 |
| Yansi | Bantu (B85) | B85 | – | – | 23 | de Filippo et al., 2011 |
| Nande | Bantu (D/J42) | D42 | – | – | 18 | Wood et al., 2005 |
| Hutu | Bantu (D/J61) | D61 | 42 | Castrì et al., 2009 | 39 | Caglià et al., 2003 |
| Hema | Bantu (E/J10) | E10 | – | – | 18 | Wood et al., 2005 |
| Ganda | Bantu (E/J15) | E15 | – | – | 26 | Wood et al., 2005 |
| Kikuyu | Bantu (E51) | E51 | 25 | Watson et al., 1996 | 42* | Wood et al., 2005 |
| Sukuma | Bantu (F21) | F21 | 32 | Knight et al., 2003; Tishkoff et al., 2007 | 32 | Knight et al., 2003; Tishkoff et al., 2007 |
| Turu | Bantu (F32) | F32 | 29 | Tishkoff et al., 2007 | 20 | Tishkoff et al., 2007 |
| Mbugwe | Bantu (F34) | F34 | – | – | 15 | Tishkoff et al., 2007 |
| Luvale | Bantu (K14) | K14 | – | – | 16 | de Filippo et al., 2011 |
| Mbunda | Bantu (K15) | K15 | – | – | 49 | de Filippo et al., 2011 |
| Ganguela | Bantu (K19) | K19 | 21 | Coelho et al., 2009 | 21 | Coelho et al., 2009 |
| Luyana | Bantu (K31) | K31 | – | – | 61** | de Filippo et al., 2011 |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|------------|----------------------|------|-------------------|-------------------------|--------------|-------------------------------|
| | | | N | Study | N | Study |
| Kwamashi | Bantu (K34) | K34 | – | – | 27 | de Filippo et al., 2011 |
| Shanjo | Bantu (K36) | K36 | – | – | 29 | de Filippo et al., 2011 |
| Fwe | Bantu (K402) | K402 | – | – | 30 | de Filippo et al., 2011 |
| Totela | Bantu (K41) | K41 | – | – | 15 | de Filippo et al., 2011 |
| Nkoya | Bantu (L62) | L62 | – | – | 15 | de Filippo et al., 2011 |
| Bisa | Bantu (M51) | M51 | 42 | de Filippo et al., 2010 | 36 | de Filippo et al., 2010; 2011 |
| Tonga | Bantu (M64) | M64 | – | – | 30 | de Filippo et al., 2011 |
| Nyanja | Bantu (N31) | N31 | 20 | Salas et al., 2002 | – | – |
| Kunda | Bantu (N42) | N42 | 36 | de Filippo et al., 2010 | 36 | de Filippo et al., 2010; 2011 |
| Nyungwe | Bantu (N43) | N43 | 20 | Salas et al., 2002 | – | – |
| Sena | Bantu (N44) | N44 | 21 | Salas et al., 2002 | – | – |
| Makonde | Bantu (P23) | P23 | 19 | Salas et al., 2002 | – | – |
| Makhuwa | Bantu (P31) | P31 | 20 | Salas et al., 2002 | – | – |
| Lomwe | Bantu (P32) | P32 | 20 | Salas et al., 2002 | – | – |
| Chwabo | Bantu (P34) | P34 | 20 | Salas et al., 2002 | – | – |
| Mbundu | Bantu (probably H21) | H21 | 44 | Plaza et al., 2004 | – | – |
| Umbundu | Bantu (R11) | R11 | 96 | Coelho et al., 2009 | 96 | Coelho et al., 2009 |
| Nkhumbi*** | Bantu (R14) | R14 | 153 | Coelho et al., 2009 | 153 | Coelho et al., 2009 |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|------------------|----------------|-----|-------------------|--|--------------|---|
| | | | N | Study | N | Study |
| Ovambos | Bantu (R21) | R21 | – | – | 28 | Forster et al., 2000 |
| Ambo | Bantu (R22) | R22 | – | – | 22 | Wood et al., 2005 |
| Herero | Bantu (R31) | R31 | – | – | 24 | Wood et al., 2005 |
| Kuvale | Bantu (R31) | R31 | 56 | Coelho et al., 2009 | 56 | Coelho et al., 2009 |
| Shona | Bantu (S10) | S10 | 77 | Salas et al., 2002; Castrì et al., 2009 | 49 | Wood et al., 2005 |
| Ndau | Bantu (S15) | S15 | 19 | Salas et al., 2002 | – | – |
| Kalanga | Bantu (S16) | S16 | – | – | 20 | de Filippo et al., 2011 |
| Sotho-Tswana | Bantu (S31–32) | S30 | – | – | 28 | Caglià et al., 2003; Wood et al., 2005 |
| Tswana | Bantu (S31) | S31 | – | – | 20 | de Filippo et al., 2011 |
| Lozi | Bantu (S34) | S34 | – | – | 94 | de Filippo et al., 2011 |
| Xhosa | Bantu (S41) | S41 | – | – | 80 | Wood et al., 2005 |
| Zulu | Bantu (S42) | S42 | – | – | 29 | Wood et al., 2005 |
| Tswa | Bantu (S51) | S51 | 19 | Salas et al., 2002 | – | – |
| Shangaan | Bantu (S53) | S53 | 22 | Salas et al., 2002 | 25 | Pereira et al., 2002 |
| Ronga | Bantu (S54) | S54 | 21 | Salas et al., 2002 | 15 | Pereira et al., 2002 |
| Chopi | Bantu (S61) | S61 | 27 | Salas et al., 2002 | – | – |
| Mbenzele Pygmies | Bantu (C10) | mbP | 57 | Destro-Bisol et al., 2004a | 52 | Coia et al., 2004 |
| Babongo Pygmies | Bantu (B42) | B42 | 45 | Quintana-Murci et al., 2008 | – | – |

| Population | subgroup | ID | Mitochondrial DNA | | Y chromosome | |
|-------------------|-----------------|------|-------------------|--|--------------|---|
| | | | N | Study | N | Study |
| Bakola Pygmies | Bantu (A801) | A801 | 88 | Tishkoff et al., 2007; Quintana-Murci et al., 2008 | 55 | Wood et al., 2005; Berniell-Lee et al., 2009 |
| Bakoya Pygmies | Bantu (B22b) | B22b | 31 | Quintana-Murci et al., 2008 | – | – |
| Biaka Pygmies | Bantu (C104) | C104 | 56 | Vigilant et al., 1991; Kivisild et al., 2006; Quintana-Murci et al., 2008 | 54 | Pritchard et al., 1999; Underhill et al., 2000; Cruciani et al., 2002; Destro-Bisol et al., 2004b; Wood et al., 2005; de Filippo et al., 2011 |

* a mixed sample of Kikuyu and Kamba

** a mixed sample of Kwangwa, Makoma, Kwandi, Kwamulonga, Mwenyi, Mbowe, Simaa, and Luyana

*** a mixed sample of Nyaneka and Nkhumbi